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3	Spot on: Using camera traps to
4	individually monitor one of the world's
5	largest lizards
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# 32 ABSTRACT

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# 34 Context

Estimating animal abundance often relies on being able to identify individuals, but this can be challenging, especially when applied to large animals which are difficult to trap and handle. Camera traps have provided a non-invasive alternative by using natural markings to individually identify animals within image data. While camera traps have been used to individually identify mammals, they are yet to be widely applied to other taxa, such as reptiles.

#### 40 Aims

41 We assessed the capacity of camera traps to provide images that allow for individual identification of 42 the world's fourth largest lizard species, the perentie (*Varanus giganteus*), and demonstrate other basic

43 morphological and behavioural data that can be gleaned from camera trap images.

### 44 *Methods*

Vertically orientated cameras were deployed at 115 sites across a 10,000km<sup>2</sup> area in north-west Australia for an average of 216 days. We used spot patterning located on the dorsal surface of perenties to identify individuals from camera trap imagery, with the assistance of freely available spot ID software. We also measured snout-to-vent length (SVL) using image analysis software, and collected image time stamp data to analyse temporal activity patterns.

### 50 Results

51 Ninety-two individuals were identified, and individuals were recorded moving distances of up to 52 1975m. Confidence in identification accuracy was generally high (91%), and estimated SVL

- 53 measurements varied by an average of 6.7% (min = 1.8%, max = 21.3%) of individual SVL averages.
- 54 Larger perentie (SVL > 45cm) were detected mostly between dawn and noon and in the late afternoon
- and early evening, whereas small perentie (SVL < 30cm) were rarely recorded in the evening.

#### 56 Conclusions

57 Camera traps can be used to individually identify large reptiles with unique markings, and can also 58 provide data on movement, morphology, and temporal activity. Accounting for uneven substrates under 59 cameras could improve the accuracy of morphological estimates. Given that camera traps struggle to 60 detect small, nocturnal reptiles, further research is required to examine whether cameras miss smaller 61 individuals in the late afternoon and evening.

### 62 Implications

63 Camera traps are increasingly being used to monitor reptile species. The ability to individually identify

64 animals provides another tool for herpetological research worldwide.

## 66 Introduction

67

Estimating species' abundance remains a key challenge within ecology and conservation biology 68 69 (Volkov et al. 2003). One obstacle is the need to distinguish between individuals within a population in 70 order to derive abundance estimates (Silver et al. 2004). For instance, mark-recapture analysis has 71 traditionally required trapping and individually marking an animal, and then using the number of re-72 traps to estimate species density and abundance (Pradel 1996). However, physically capturing animals 73 is not always practical (Sanecki and Green 2005) nor desirable (De Bondi et al. 2010), and therefore 74 non-invasive methods that use unique morphological features to passively identify individuals are 75 sometimes more appropriate (Brooks et al. 2010; Silver et al. 2004).

76

77 Camera traps provide a means of collecting image-based data that can be used to distinguish marked 78 (scaring, natural markings) individuals from one another (Foster and Harmsen 2012; Higashide et al. 79 2012), and are increasingly used to estimate species density/abundance using mark-recapture analysis 80 (Burton et al. 2015). So far, studies that have used camera trap imagery to identify individual animals 81 and estimate density/abundance have been almost entirely limited to mammalian taxa (Burton et al. 82 2015), such as tigers (Panthera tigris)(Jhala et al. 2011), bobcats (Lynx rufus) (Alonso et al. 2015), 83 leopards (Panthera pardus) (Rostro-García et al. 2018) and puma (Puma concolor)(Alexander and 84 Gese 2018). Despite increasing recognition that camera traps provide a useful method with which to 85 survey reptiles (Molyneux et al. 2018; Richardson et al. 2018; Welbourne et al. 2015; Welbourne et al. 86 2017), few studies have used camera trap imagery to identify individuals within a reptile species 87 (Bennett and Clements 2014; Welbourne 2013), although other studies have identified individuals using 88 manually operated cameras (Kellner et al. 2017; Moro and MacAulay 2014; Treilibs et al. 2016). 89

90 Species within the predatory lizard genus Varanus are notoriously difficult to trap and handle (Green 91 and King 1978; Jessop et al. 2006), given they can weigh >90kg and measure three metres in length 92 (Jessop et al. 2006), and yet the need to monitor these species has never been greater. Varanids 93 (Varanus.spp) are widely distributed across Australia, Asia and Africa, and often fulfil the role of a 94 mesopredator or apex predator (King and Green 1999). Multiple species of varanids are threatened from 95 exploitation by humans (meat, leather) (Shine and Harlow 1998), the illegal pet trade (Ruxmoore and 96 Groombridge 1990), habitat loss (Gibbons et al. 2000) and introduced species (Shine 2010). The 97 removal of these important predators from ecosystems can have cascading impacts on a range of other 98 species (Doody et al. 2006; Read and Scoleri 2015; Sutherland et al. 2010), highlighting the need for a 99 more accurate understanding of their populations.

100

Here, we use camera trap imagery to identify individuals of the world's fourth largest species of lizard,
the Perentie (*Varanus giganteus*). Perentie measure in excess of 2 m long, weigh up 20 kg, and are

103 potentially venomous (Fry et al. 2006; Wilson and Swan 2017), however little is known about the 104 ecology of this species. The use of non-capture methods to study perentie are therefore particularly 105 desirable, especially given trapping these animals likely poses risks to both the animal and the handler. 106 Further, their large size means that perentie tend to evade capture/detection using traditional reptile 107 survey techniques such as pitfall trapping (Richardson et al. 2018). The scales of perentie are covered 108 with lateral bands of large yellow spots that are unique to the individual (Moro and MacAulay 2014). 109 In this study, we investigate the feasibility of individually identifying perentie at a landscape-scale using 110 remote sensing cameras. We also use scaled camera trap images to estimate individual body lengths, 111 and time stamp data to observe temporal trends — both techniques which have never been used on a 112 varanid species and are very rarely applied to reptiles in general.

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### 115 Methods

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- 117 *Study site*
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This study was conducted across four cattle stations within the Pilbara bioregion in north-west Australia. This area also encompassed the Karayarra and Nyamal indigenous language groups (Fig B1).
Vegetation within this area is mostly dominated by *Triodia* grasslands (McKenzie *et al.* 2009). Geology in the Pilbara is characterised by largely flat sand plains and granite outcrops (Doughty *et al.* 2011; Withers 2000). Average annual rainfall within the study area is 339.5mm (Indee station) (BOM 2020).

124 Survey design

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We used a whole-of-landscape experimental design, in which multiple sample sites are embedded 126 127 within a heterogeneous area (Fahrig 2003), commonly termed a landscape (Bennett et al. 2006). Twenty 128 three study landscapes were selected using ArcMap 10.3 (ESRI 2011). In our study, landscapes were 129 circular with a diameter of 1 km (area =75 ha), and were selected to contain patches of rocky habitat dispersed within a matrix of spinifex grasslands. We deployed five camera sites within each landscape 130 131 (115 sites in total), each separated by > 200 meters. All study sites were placed within rocky outcrops, 132 as this habitat is known to be utilized by perenties and other species (e.g., northern quolls, *Dasyurus* 133 *hallucatus*) that were the focus of the broader research program within which this study was embedded 134 (Menkhorst and Knight 2001; Wilson and Swan 2017). 135

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Figure B1 – Twenty three study landscapes spread across four cattle stations in the Pilbara bioregion
 — Pippingarra, Indee, Yandeyarra and Mallina. Grey lines represent cattle station boundaries. Five
 downward facing camera sites were deployed at each study landscape.

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146 We used Reconyx<sup>™</sup> PC900 Hyperfire covert cameras (Reconyx, Wisconsin, USA). These cameras use 147 an infrared flash. Cameras were attached to a wooden tree stake 1.5m above the ground and were 148 positioned to face downward, with the camera lens focused directly at the ground surface using a 149 bookshelf bracket (Appendix 1). A PVC canister containing approximately 150g of bait (canned fish) 150 was attached to the bottom of the tree stake supporting the camera. Twelve landscapes were sampled 151 between August 2017 and April 2018. The remaining 11 landscapes were sampled between August 152 2018 and April 2019. Average camera deployment time was 216 (min =151, max = 245) days at each 153 site. Bait was replenished twice at all sites during this period after roughly 70 and 140 days of camera 154 deployment. At yandeyarra station, camera placement was partly guided by knowledge from Indigenous rangers as part of the Greening Australia ranger program. Cameras were set to high sensitivity with no 155 156 delays between triggers, and five images were taken at one second intervals per trigger. We defined an 157 independent detection event as consecutive triggers of the same individual separated by greater than 15 158 minutes.

161

# 160 Individual identification

We used spot patterning located on the dorsal surface of perentie to identify individuals from camera trap imagery (Figure B2). To do this, we first catalogued images from each detection event that best represented an individual's unique spot pattern. Only images with at least 30% of the animal's dorsal surface patterning visible were catalogued for analysis. We found that attempting to identify animals in images with less than 30% of the animal's dorsal surface viable was generally unreliable, as fewer spots markings are available to distinguish between animals.

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169 We then entered one or more images from each catalogued detection event into I3S (Den Hartog and 170 Reijns 2016) —a freely available spot ID program that has previously been used to identify individuals 171 in other spotted species such as spotted eagle rays (Aetobatus narinari) (Cerutti-Pereyra et al. 2018), 172 Italian crested newts (Triturus carnifex) (Sannolo et al. 2016) and whale sharks (Rhincodon typus) 173 (Rowat et al. 2009). I3S automatically compares a target individual's spot pattern to a library of other 174 individual spot patterns that have been pre-entered into the system by an observer. To do this, the I3S 175 uses a two-dimensional linear algorithm, which compares spot coordinates, as well as spot shape and 176 size between patterns. I3S then ranks which patterns within the existing library are most similar to the 177 target spot pattern in the form of a pattern difference score, where lower pattern difference scores 178 correspond to increasing pattern similarity (Cerutti-Pereyra et al. 2018; Speed et al. 2007). Once 179 patterns had been ranked, at least two observers manually confirmed if the pattern suggested by the I3S 180 program matched the target spot pattern. If no match could be found, the target individual was assigned 181 a reference code and entered into the data base as a new animal.

182

183 To gauge observer confidence in individual identification accuracy, we marked each detection event 184 with a confidence rating using an adapted identification protocol outlined in (Hohnen et al. 2013). 185 Confidence rating were derived from the number of unique markings/features that could used to identify 186 an animal. An observer had 'high' confidence that an individual has been accurately identified if at least 187 five unique markings/features could be used to identify an animal. An observer had 'medium' 188 confidence that an individual has been accurately identified if between two and five markings/features 189 could be used to identify an animal. An observer had 'low' confidence that an individual has been accurately identified if only one marking/feature could be used to identify an animal. For individuals 190 191 that were recorded at more than one site, we measured distance travelled using the measure function in 192 ArcMap 10.3 (ESRI 2011).

193

We used an analytical Bayesian approach with a vague gamma prior to summarise the number of detections per perentie, as well as the number of perenties per site and per landscape. This approach

- 196 produces upper and lower 95% credible intervals, which are comparable to confidence intervals
- 197 produced when using a frequentist approach.
- 198 We used credible intervals because standards estimates of mean error (ie standard deviation) in count
- 199 data where means are close to zero can produce nonsensical outputs, such as negative counts.





Figure B2 – An example of the spot patterning used to individually identify perentie (*V. giganteus*) within the Pilbara bioregion. The three images shown are of the same animal, and were taken over a period of 57 days (23/10/2017 - 19/12/2017) at three different camera sites. Red circles represent specific spots that were used to identify this animals.

### 233 Length measurement

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235 We measured the snout-to-estimated vent length (SVL) of perentie individuals in camera trap images using the computer program imageJ (Abramoff et al. 2004). Because the vent was not visible from the 236 237 dorsal surface, we consistently estimated the vent location as being at the base of the hind legs in the 238 centre of the animal's body, based on morphometric measurement protocol used by Thompson and 239 Withers (1997). SVL measurements followed the animal's spine from the snout to the vent to increase 240 measurement accuracy. ImageJ allows users to measure distances within imagery by calculating a pixel-241 distance (mm) ratio from an object with a known length (scale) within an image. Once a pixel-distance 242 ratio has been determined, objects with an unknown length can be measured. Here, we used brackets 243 located at the base of tree stakes supporting cameras as the scale (40 mm). To test the accuracy of using 244 image-j to measure animal sizes from camera trap imagery, we conducted a pilot test using Reconyx 245 PC900 Hyperfire covert cameras and target objects with known dimensions. We found imageJ 246 measurements varied from actual measurements (using a measuring tape) by an average of < 3%. To 247 visualize perentie SVL data, we created histograms of SVL using ggplot in R version 3.5.3 (R Core 248 Team 2020; Wickham and Wickham 2007). One SVL measurement was taken for each independent 249 detection. When an individual was detected multiple times, and thus multiple SVL measurements were 250 taken, we used the mean measurement.

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#### 252 Temporal activity

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To further illustrate the range of data that can be collected for lizards using camera traps, we used time stamp data from detection images to fit non-parametric kernel density curves using the '*overlap*' package in R (Ridout and Linkie 2009), giving a probability density distribution of perentie activity patterns .This method is commonly used to quantify species temporal activity patterns within ecological studies (Allen *et al.* 2018; Azevedo *et al.* 2018; Fancourt *et al.* 2015; Lashley *et al.* 2018). We used a default smoothing parameter of 0.8 in our analysis, as recommended by Ridout and Linkie (2009) for small sample sizes.

261

262 Perenties being ectotherms, are reliant on external sources of heat to maintain a constant body 263 temperature, which can determine when they are most likely to be active throughout the day. Because 264 larger lizards are generally able to retain body heat longer than smaller lizards (Stevenson 1985), we 265 expected body size to impact perentie temporal activity. To account for these differences, we analysed perentie temporal activity using three sizes classes; large (SVL >45 cm), medium (SVL = 30 to 45cm) 266 and small (SVL < 30 cm). We determined sizes classes based on the distribution of SVL measurements 267 268 collected in this study. To elucidate if perenties of different size classes were detected more frequently 269 at different times, we measured overlap in temporal activity between large, medium, and small

- 270 individuals, using an overlap coefficient ( $\Delta$ ). Overlap coefficients are the product of a function which 271 describes the distance between two temporal activity density distributions, and can range from zero (no 272 overlap) to one (complete overlap) (Ridout and Linkie 2009). We used the package overlap' (Meredith 273 and Ridout 2014) in R version 3.5.3 (R Core Team 2020) to calculate temporal overlap between 274 perentie size classes, which produces three overlap coefficients for each temporal comparison. The most 275 appropriate coefficient for estimating overlap is dependent on sample size (Ridout and Linkie 2009). 276 We used the overlap coefficient  $\Delta_1$  as it is most appropriate for small datasets (Ridout and Linkie 2009). 277 Overlap precision was measured using confidence intervals generated from 500 random bootstrapped 278 samples after accounting for bootstrap bias (Meredith and Ridout 2014).
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## 280 **Results**

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282 We recorded 190 detections of perentie over 28,840 trap nights. Other varanid species detected included 283 Varanus panoptes, V. pilbarensis, V. gouldii and V. acanthurus. Perentie were detected at 91.3% of 284 landscapes and 48.6% of sites. Seventy-three percent of detection events were suitable for individual 285 identification, and from these, we identified a total of 92 individuals using spot pattern analysis. 286 Confidence in identification accuracy was generally high (91%) or medium (7%). Thirty-one (33.7%) 287 of individuals were recaptured at least once. The number of detections per individual ranged from one to five ( $\bar{x} = 1.55$ , lower credible interval = 1.34, upper credible interval = 1.82), and the number of 288 289 individuals detected within a single landscape ranged from zero to twenty ( $\bar{x} = 3.99$ , lower = 3.22, upper 290 = 4.86). The number of individuals per site ranged from one to 13 ( $\bar{x}$  = 0.80, lower = 0.64, upper = 0.97) 291 (Figure B3), and thirteen individuals were detected at more than one site (Figure B4,B5). The average 292 total distance travelled between sites where the same individual was detected was 887 metres (sd = 632293 m), ranging from 224 m to 1975 m (Figure B5). Average estimated SVL length was 37.22 cm (sd = 294 12.85 cm), ranging from 10.29 cm to 73.30 cm (Figure B6). When an individual was detected multiple 295 times, length measurements varied by an average of 2.46cm (min = 0.64, max = 7.86), or 6.7% (min = 296 1.8%, max = 21.3%) of individual SVL averages (Appendix 2). SVL increased by 5cm and 7cm for 297 two individuals that were detected 123 and 124 days respectively after they were initially detected, 298 potentially as a result of growth. We found limited evidence of growth otherwise. Twenty-nine percent 299 of individuals measured were classified as small (<30cm), 44% were classified as medium (30–45cm) 300 and 29% were classified as large (>45cm).

301

302 Temporal data showed on average perentie were most active between dawn and noon, and around dusk

303 (Figure B7). Overlap in activity patterns was highest between medium and small individuals ( $\Delta = 0.80$ ,

lower CI = 0.63, upper CI = 0.89) and lowest between large and small individuals ( $\Delta$  = 0.67, lower CI

305 = 0.43, upper CI = 0.81). Overlap between large and medium individuals was 0.80 (lower CI = 0.55,

306 upper CI = 0.85). Large individuals were most active between dawn and noon but also active at dusk 307 and during the night, whereas small individuals were almost exclusively active between dawn and noon. 308

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309

### 311 **Discussion**

312

Camera trap imagery is commonly used to identify individuals within mammal populations (Burton et 313 314 al. 2015), but few have been used to individually identify lizards (Bennett and Clements 2014; 315 Welbourne 2013). In this study, we used camera traps to identify 92 individual perenties across 23 316 landscapes within the Pilbara bioregion, Australia. We found a large proportion of independent 317 detections were suitable for individual identification, and observer confidence in identification accuracy 318 was high, suggesting camera traps may offer a practical alternative to live trapping when monitoring 319 some large reptile species. We also highlight that other useful data, such as movement data, body length, 320 and temporal activity patterns can be derived from the same imagery. This research highlights the 321 significant potential of camera traps for population monitoring of large reptiles across the world.

322

323 The capacity to individually identify animals from camera trap data unlocks the use of a variety of 324 statistical analyses that can be used to estimate population densities, as well as survival probabilities 325 and population spatial dynamics. For example, mark-recapture models rely on individual identification 326 to estimate abundance (Pradel 1996). We show that collecting the data necessary to perform such 327 analyses on large lizards is possible through the use of remote sensing cameras. However, it is important 328 to note that in addition to the ability to identify individuals, other data assumptions must be met in order 329 to reliably estimate abundance or density using mark-recapture methods (Pradel 1996). For example, 330 for most mark-recapture models, accuracy is highest when populations are closed (Kendall 1999); that 331 is no deaths, births, immigration or emigration occurs across sampling periods. Whilst it is possible 332 individuals left or entered the study population during sampling periods in our study, the likelihood of 333 this occurring can be reduced by shortening the sampling period, or using mark-recaptures models that 334 account for open populations (Schwarz and Arnason 1996). Low recapture rates as a result of poor 335 capture efficiency can also impact the reliability of mark-recapture estimates (Morton 1982; Pollock 336 1980). In our study, we found a third of individuals were recaptured at least once, and some individuals 337 were recaptured up to four times. This result suggests capture efficiency may be adequate for mark-338 recapture methods to be applied on data collected in this study, however it would be possible to increase 339 capture efficiency by increasing the number of cameras deployed per landscape.

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342 Perentie were detected across most study landscapes, and some individuals were detected across 343 multiple landscapes, moving distances of almost 2km. Whilst the spatial ecology of perentie remains 344 poorly understood, research focused on other varanids suggests movements of this distance are not 345 unusual (Green and King 1978; Guarino 2002). We also found a number of individuals were recorded 346 visiting the same camera sites repeatedly (up to five times), sometimes over a period of months. Whilst 347 we may only speculate where these individuals travelled in the time between detections, habitual visits 348 to the same site could indicate some individuals may occupy a home range like other varanids (Lei and 349 Booth 2018).

350

351 One disadvantage associated with using non-capture related techniques to survey wildlife is the lack of 352 ability to collect morphological data. In this study, we were able to approximate perentie SVL by scaling 353 images to a known distance. SVL length varied substantially between individuals suggesting camera 354 traps were able to detect a range of demographics ranging from juveniles (SVL = 10.3 cm) to larger 355 adults (SVL = 73.3 cm). Average SVL length was 37.22 cm (min = 10.3 cm, max = 73.3 cm), which fits 356 within the bounds of previous measurements that were recorded in situ on perentie in Western Australia. For example Thompson and Withers (1997) found average SVL length from 25 captured perentie was 357 358 44.2 cm (sd = 12.4 cm), ranging from 15.9 cm - 66 cm. Similarly King *et al.* (1989) found SVL length 359 of captured perentie on Barrow Island ranged from 23 cm - 88 cm (no average available). Whilst 360 estimating body length using scaled imagery is an accepted method within wildlife research (Weinstein 361 2018), in our study, uneven ground surfaces on top of which lizards were measured has the potential to 362 distort estimates. Despite this, we found measurement consistency was relatively high (average 363 variation from mean individual SVL = 6.7%). This result suggests that, although SVL measurements 364 using camera trap imagery are obviously less accurate than measurements obtained with animals in 365 hand, they may be able to provide reasonable estimates of animal size that could potentially be used to 366 infer other individual attributes such as age and sex (although see Smith et al. 2007). A practical means of improving length estimates using camera traps in future studies may be to install a flat base plate 367 underneath cameras (following Welbourne 2013; Welbourne et al. 2015), so that variation in the surface 368 369 on which animals are measured is removed.

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371 Behavioural data can be important to species conservation (French et al. 2019; Tyne et al. 2017), yet 372 accessing this data usually involves tracking an animal using expensive satellite telemetry equipment 373 (Bastille-Rousseau et al. 2018; Hertel et al. 2019), or investing large quantities of time manually 374 recording animal activity in the field (Brieger et al. 2017; Li et al. 2017). We used time stamp data 375 imprinted on camera trap imagery to record perentie diel activity. Temporal activity varied between 376 perentie size classes, however overlap between perentie size class combinations was mostly similar 377  $(\Delta 0.67 - \Delta 0.80)$ . The most notable differences in temporal activity were observed in temporal activity 378 peaks. For example, activity peaks for large and medium sized individuals were spread across much of 379 the 24-hour diel period, occurring at mid-morning, dusk and midnight, whereas activity for small 380 individuals was mostly confined to mid-morning and early afternoon. Observed temporal differences 381 could be related to individual physiology, where individuals with larger body mass are able to 382 thermoregulate more efficiently than individuals with smaller body mass, and thus stay active longer 383 (Garrick 2008). Alternatively, smaller individuals may avoid being active around dusk and at night to 384 reduce their likelihood of encountering potential predators such as feral cats (*Felix catus*) and dingoes 385 (*Canis dingo*) which are generally most active at these times within the study area (GéCzy 2009; 386 Hernandez-Santin et al. 2016; Johnson 1976). Whilst some varanids are known to predate on smaller members of their own species (Polis and Myers 1985), our results do not suggest smaller perenties are 387 388 temporally avoiding larger perenties. This is because peak activity of large and small perenties overlaps 389 considerably around 11:00am. Further, smaller perenties are not active at any time larger perenties are 390 not active.

391

392 Another explanation is that cameras fail to detect smaller lizards at night due to a lack of thermal 393 contrast. This is consistent with the findings of Richardson et al. (2018), who found that camera traps 394 failed to detect small, nocturnal reptiles. Passive infrared triggered cameras, as used in this experiment, 395 will detect an animal when a difference between the animals surface temperature and the background 396 temperature is detected, (Welbourne et al. 2016), with optimal conditions for detection occurring when 397 the temperature difference is at least 2.7° C (Meek et al. 2012). When the background temperature is 398 similar to an animals surface temperature, the capacity of PIR cameras to detect the animal may be 399 reduced (Rovero et al. 2013). On the basis of these facts and the laws of thermodynamics, the surface 400 temperature of smaller perentie individuals maybe more similar to the background temperature — 401 derived from the red granite on top of which cameras were positioned — because they have a smaller 402 body mass, and therefore maybe less likely to be detected by PIR sensors. Contrast between the 403 background temperature and surface temperature of smaller perenties may be larger in the day because 404 animals are better able to thermoregulate by utilizing microhabitats that are cooler than the rest of their 405 environment (Sears et al. 2016). To increase thermal contrast when using PIR cameras to detect lizards 406 Welbourne (2013) successfully augmented background temperature using cork tiles, making the lizards 407 more visible to sensors. Whilst implementing a similar technique would have been desirable in this 408 study, unfortunately it was logistically not possible.

409

Recent improvements in remote sensing technologies have created opportunities for wildlife research to utilize non-invasive labour efficient techniques instead of methods that involve the live trapping of animals. Importantly, we recognise that although many large species of lizards bare natural markings that could potentially be used to tell animals apart (Chen *et al.* 2013; King and Horner 1987; Rodda *et al.* 1988), consistent individual identification may not be feasible for all species, especially those which lack spot or line based scale patterning. One potential objective of future research could be to explore

416	the practicality of using camera traps to individually identify animals from other lizard taxa that exhibit
417	visible scale patterning, such as species with the Agamidae and Scincidae families (Welbourne 2013).
418	Another objective could be to trial alternative camera types and settings that may be better suited to
419	detecting smaller species, whilst still providing high resolution imagery. Preliminary research in this
420	area suggests cameras programmed to take images at standard intervals capture more images of
421	squamates than cameras triggered by sensors, and cameras with a manually adjusted focus collect
422	images better suited to identifying smaller lizards than cameras which use default focus settings
423	(Welbourne et al. 2019). In this study, we demonstrate how camera traps can be used to detect
424	Australia's largest species of lizard and provide imagery that can be used to differentiate individuals as
425	well as observe temporal behaviour. Overall the findings of our study provide a promising indication
426	of the potential uses for camera traps in studying large lizards and herpetological research in general.
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437 Figure B3 – Boxplots depicting the total number of perentie (*Varanus giganteus*) individuals detected
438 at study sites grouped within the 23 study landscapes within the Pilbara bioregion in Western Australia.
439 Five study sites were within each study landscape.

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Figure B4 – Perentie (Varanus giganteus) individuals detected at multiple study sites within the Pilbara bioregion in Western Australia. a.) Individual 02\_YA\_04 was detected at site Q0704 on 30/09/2018 before moving 508 metres to site Q0701 on 30/10/2018 and then 431 metres to site Q0703 on 05/01/2019. b.) Individual 01\_MA\_13 was detected at site Q6603 on 25/09/2017 before moving 1573 metres to site Q3205 on the 8/11/2017. Individual 01\_MA\_09 was detected at site Q3202 on 15/01/2018 before moving 1975 metres to site Q6601 on 10/03/2018. c.) Individual 02\_YA\_04 was detected at site Q0704 on 30/09/2018 before moving 508 metres to site Q0701 on 30/10/2018 and then 431 metres to site Q0703 on 05/01/2019. d.) Individual 01\_ID\_09 was detected at site Q2201 on 23/10/2017 before being detected 201 metres away at site Q2202 on 24/10/2017 and then 1245 metres away at site Q1905 on 19/12/2017.





484 Figure B5 – Histogram depicting total distances travelled (m) by the 13 Perentie (*V..giganteus*)
485 individuals that were detected at more than one study landscape within the Pilbara bioregion .





493 Figure B6 – Histogram depicting the averaged snout to estimated vent length (cm) of 73 Perentie (V.
494 *giganteus*) individuals that were measured from camera trap image using freely available image analysis
495 software.



Figure B7 – Non-parametric kernel density curves depicting temporal activity for Perentie (Varanus giganteus) in the Pilbara bioregion recorded using camera traps. Grey sections represent overlap in temporal activity. Large individuals were individuals with an average SVL exceeding 45cm. Medium individuals were individuals with an average SVL between 30 cm and 45 cm. Small individuals were individuals with an average SVL less than 30cm. A.) Averaged temporal activity for all individuals B.) Overlap in temporal activity for large and medium individuals C.) Overlap in temporal activity for medium and small individuals D.) Overlap in temporal activity for large and small individuals.

- 520 Appendices



523 Figure BS1– Schematic of camera trap design used to detect and identify Perentie (*V. giganteus*)
524 individuals.



531 Figure BS2 – Boxplot depicting variation in the snout to estimated vent length (cm) of 15 Perentie (V.
533 giganteus) individuals that were measured from camera trap image using freely available image analysis

- 534 software

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574

- Abramoff, M. D., Magalhaes, P. J., and Ram, S. J. (2004). Image Processing with ImageJ. *Biophotonics International* 11, 36-42.
- 579 Alexander, P. D. and Gese, E. M. (2018). Identifying individual cougars (Puma concolor) in remote 580 camera images–implications for population estimates. *Wildlife research* **45**, 274-281.
- 581

Allen, M. L., Peterson, B., and Krofel, M. (2018). No respect for apex carnivores: distribution and activity patterns of honey badgers in the Serengeti. *Mammalian biology* **89**, 90-94.

584

Alonso, R. S., McClintock, B. T., Lyren, L. M., Boydston, E. E., and Crooks, K. R. (2015). Markrecapture and mark-resight methods for estimating abundance with remote cameras: a carnivore case study. *PloS one* **10**, e0123032.

588

Azevedo, F., Lemos, F., Freitas-Junior, M., Rocha, D., and Azevedo, F. (2018). Puma activity patterns
 and temporal overlap with prey in a human-modified landscape at Southeastern Brazil. *Journal of Zoology* 305, 246-255.

592

596

602

604

608

- Bastille-Rousseau, G., Wall, J., Douglas-Hamilton, I., and Wittemyer, G. (2018). Optimizing the
  positioning of wildlife crossing structures using GPS telemetry. *Journal of Applied Ecology* 55, 20552063.
- Bennett, A. F., Radford, J. Q., and Haslem, A. (2006). Properties of land mosaics: implications for
  nature conservation in agricultural environments. *Biological conservation* 133, 250-264.
- 600 Bennett, D. and Clements, T. (2014). The use of passive infrared camera trapping systems in the study 601 of frugivorous monitor lizards. *Biawak* **8**, 19-30.
- 603 BOM (2020). Bureau of Meteorology.)
- Brieger, F., Hagen, R., Kröschel, M., Hartig, F., Petersen, I., Ortmann, S., and Suchant, R. (2017). Do
  roe deer react to wildlife warning reflectors? A test combining a controlled experiment with field
  observations. *European Journal of Wildlife Research* 63, 72.
- Brooks, K., Rowat, D., Pierce, S. J., Jouannet, D., and Vely, M. (2010). Seeing spots: photoidentification as a regional tool for whale shark identification. *Western Indian Ocean Journal of Marine Science* 9, 185-194.
- Burton, A. C., Neilson, E., Moreira, D., Ladle, A., Steenweg, R., Fisher, J. T., Bayne, E., and Boutin,
  S. (2015). Wildlife camera trapping: a review and recommendations for linking surveys to ecological
  processes. *Journal of Applied Ecology* 52, 675-685.
- 616
  617 Cerutti-Pereyra, F., Bassos-Hull, K., Arvizu-Torres, X., Wilkinson, K., García-Carrillo, I., Perez618 Jimenez, J., and Hueter, R. (2018). Observations of spotted eagle rays (*Aetobatus narinari*) in the
  619 Mexican Caribbean using photo-ID. *Environmental biology of fishes* 101, 237-244.
- 620 621 Chen, I.-P., Symonds, M. R., Melville, J., and Stuart-Fox, D. (2013). Fac
- Chen, I.-P., Symonds, M. R., Melville, J., and Stuart-Fox, D. (2013). Factors shaping the evolution of
  colour patterns in Australian agamid lizards (*Agamidae*): a comparative study. *Biological Journal of the Linnean Society* 109, 101-112.
- De Bondi, N., White, J. G., Stevens, M., and Cooke, R. (2010). A comparison of the effectiveness of camera trapping and live trapping for sampling terrestrial small-mammal communities. *Wildlife research* **37**, 456-465.
- 629 Den Hartog, J. and Reijns, R. (2016). Interactive Individual Identification Software (I3S).)
- 630

- Doody, J., Green, B., Sims, R., Rhind, D., West, P., and Steer, D. (2006). Indirect impacts of invasive
  cane toads (*Bufo marinus*) on nest predation in pig-nosed turtles (*Carettochelys insculpta*). Wildlife *research* 33, 349-354.
- 634

641

644

651

655

659

662

664

670

673

680

- Doughty, P., Rolfe, J. K., Burbidge, A. H., Pearson, D. J., and Kendrick, P. G. (2011). Herpetological
  assemblages of the Pilbara biogeographic region, Western Australia: ecological associations,
  biogeographic patterns and conservation. *Records of the Western Australian Museum, Supplement* 78,
  315-341.
- 640 ESRI (2011). ArcGIS Desktop: Release 10.)
- Fahrig, L. (2003). Effects of habitat fragmentation on biodiversity. *Annual review of ecology, evolution, and systematics* 34, 487-515.
- Fancourt, B. A., Hawkins, C. E., Cameron, E. Z., Jones, M. E., and Nicol, S. C. (2015). Devil declines
  and catastrophic cascades: is mesopredator release of feral cats inhibiting recovery of the eastern quoll? *PloS one* 10, e0119303.
- Foster, R. J. and Harmsen, B. J. (2012). A critique of density estimation from camera-trap data. *The Journal of wildlife management* 76, 224-236.
- French, R. K., Muller, C. G., Chilvers, B. L., and Battley, P. F. (2019). Behavioural consequences of
  human disturbance on subantarctic Yellow-eyed Penguins, *Megadyptes antipodes*. *Bird Conservation International* 29, 277-290.
- Fry, B. G., Vidal, N., Norman, J. A., Vonk, F. J., Scheib, H., Ramjan, S. R., Kuruppu, S., Fung, K.,
  Hedges, S. B., and Richardson, M. K. (2006). Early evolution of the venom system in lizards and snakes. *Nature* 439, 584-588.
- Garrick, D. (2008). Body surface temperature and length in relation to the thermal biology of lizards. *Bioscience Horizons* 1, 136-142.
- 663 GéCzy, C. (2009). Cannibalism in captive Varanus timorensis. Biawak 3, 61-63.
- Gibbons, J. W., Scott, D. E., Ryan, T. J., Buhlmann, K. A., Tuberville, T. D., Metts, B. S., Greene, J.
  L., Mills, T., Leiden, Y., and Poppy, S. (2000). The Global Decline of Reptiles, Déjà Vu Amphibians:
  Reptile species are declining on a global scale. Six significant threats to reptile populations are habitat
  loss and degradation, introduced invasive species, environmental pollution, disease, unsustainable use,
  and global climate change. *BioScience* 50, 653-666.
- Green, B. and King, D. (1978). Home range and activity patterns of the sand goanna, *Varanus gouldii*(Reptilia: *Varanidae*). *Wildlife research* 5, 417-424.
- 674 Guarino, F. (2002). Spatial ecology of a large carnivorous lizard, *Varanus varius* (Squamata: 675 *Varanidae*). *Journal of Zoology* **258**, 449-457.
- 676
  677 Hernandez-Santin, L., Goldizen, A. W., and Fisher, D. O. (2016). Introduced predators and habitat
  678 structure influence range contraction of an endangered native predator, the northern quoll. *Biological*679 *conservation* 203, 160-167.
- Hertel, A. G., Leclerc, M., Warren, D., Pelletier, F., Zedrosser, A., and Mueller, T. (2019). Don't poke
  the bear: using tracking data to quantify behavioural syndromes in elusive wildlife. *Animal behaviour*147, 91-104.
- Higashide, D., Miura, S., and Miguchi, H. (2012). Are chest marks unique to A siatic black bear

- 686 individuals? *Journal of Zoology* **288**, 199-206.
- Hohnen, R., Ashby, J., Tuft, K., and McGregor, H. (2013). Individual identification of northern quolls
  (*Dasyurus hallucatus*) using remote cameras. *Australian Mammalogy* 35, 131-135. doi: 10.1071/am12015.
- Jessop, T. S., Madsen, T., Sumner, J., Rudiharto, H., Phillips, J. A., and Ciofi, C. (2006). Maximum
  body size among insular Komodo dragon populations covaries with large prey density. *Oikos* 112, 422429.
- 695

691

- Jhala, Y., Qureshi, Q., and Gopal, R. (2011). Can the abundance of tigers be assessed from their signs?
   *Journal of Applied Ecology* 48, 14-24.
- 698

711

714

718

- Johnson, C. R. (1976). Some behavioural observations on wild and captive sand monitors, *Varanus gouldii* (Sauria: *Varanidae*). *Zoological Journal of the Linnean Society* 59, 377-380.
- Kellner, C. J., Lawson, G. R., Tomke, S. A., and Noble, J. H. (2017). Computer-aided individual
  identification of Sceloporus consobrinus based on patterns of head scalation. *Herpetol. Rev* 48, 766769.
- Kendall, W. L. (1999). Robustness of closed capture–recapture methods to violations of the closure assumption. *Ecology* 80, 2517-2525.
- King, D. and Green, B. (1999) 'Goannas: the biology of varanid lizards.' (UNSW Press: Sydney, 2052,
   Australia.)
- King, D., Green, B., and Butler, H. (1989). The Activity Pattern, Temperature Regulation and Diet of
  Varanus-Giganteus on Barrow-Island, Western-Australia. *Wildlife research* 16, 41-47.
- King, M. and Horner, P. (1987). A new species of monitor (Reptilia: Platynota) from northern Australia
  and a note on the status of Varanus acanthurus insulanicus Mertens. *Beagle: Records of the Museums and Art Galleries of the Northern Territory, The* 4, 73-77.
- Lashley, M. A., Cove, M. V., Chitwood, M. C., Penido, G., Gardner, B., DePerno, C. S., and Moorman,
  C. E. (2018). Estimating wildlife activity curves: comparison of methods and sample size. *Scientific reports* 8, 4173.
- Lei, J. and Booth, D. T. (2018). Intraspecific variation in space use of a coastal population of lace
  monitors (*Varanus varius*). *Australian Journal of Zoology* 65, 398-407.
- Li, D., Liu, Y., Sun, X., Lloyd, H., Zhu, S., Zhang, S., Wan, D., and Zhang, Z. (2017). Habitatdependent changes in vigilance behaviour of Red-crowned Crane influenced by wildlife tourism. *Scientific reports* 7, 16614.
- McKenzie, N., Van Leeuwen, S., and Pinder, A. (2009). Introduction to the Pilbara biodiversity survey,
   2002–2007. *Records of the Western Australian Museum, Supplement* 78, 3-89.
- Meek, P. D., Fleming, P., and Ballard, G. (2012) 'An introduction to camera trapping for wildlife
  surveys in Australia.' (Invasive Animals Cooperative Research Centre Canberra, Australia.)
- Menkhorst, P. and Knight, F. (2001) 'Field guide to the mammals of Australia.' (Oxford University
  Press: New York.)
- 739 Meredith, M. and Ridout, M. (2014). Overview of the overlap package. In 'R Proj' pp. 1-9.)
- 740

738

- Molyneux, J., Pavey, C., James, A., and Carthew, S. (2018). The efficacy of monitoring techniques for detecting small mammals and reptiles in arid environments. *Wildlife research* **44**, 534-545.
- 743

755

758

761

768

774

Moro, D. and MacAulay, I. (2014). Computer-aided pattern recognition of large reptiles as a
noninvasive application to identify individuals. *Journal of applied animal welfare science* 17, 125-135.

- Morton, A. (1982). The effects of marking and capture on recapture frequencies of butterflies. *Oecologia* 53, 105-110.
- Polis, G. A. and Myers, C. A. (1985). A survey of intraspecific predation among reptiles and amphibians. *Journal of Herpetology* **19**, 99-107.
- Pollock, K. H. (1980) 'Capture-recapture models: a review of current methods, assumptions and
  experimental design.' (Citeseer: North Carolina 27650, USA.)
- Pradel, R. (1996). Utilization of capture-mark-recapture for the study of recruitment and population growth rate. *Biometrics* **52**, 703-709.
- R Core Team, R. (2020). R: A language and environment for statistical computing. *R Foundation for statistical computing, Vienna.*
- Read, J. L. and Scoleri, V. (2015). Ecological implications of reptile mesopredator release in arid South
  Australia. *Journal of Herpetology* 49, 64-69.
- Richardson, E., Nimmo, D. G., Avitabile, S., Tworkowski, L., Watson, S. J., Welbourne, D., and
  Leonard, S. W. (2018). Camera traps and pitfalls: an evaluation of two methods for surveying reptiles
  in a semiarid ecosystem. *Wildlife research* 44, 637-647.
- Ridout, M. S. and Linkie, M. (2009). Estimating overlap of daily activity patterns from camera trap
  data. *Journal of Agricultural, Biological, and Environmental Statistics* 14, 322-337.
- Rodda, G. H., Bock, B. C., Burghardt, G. M., and Rand, A. S. (1988). Techniques for identifying
  individual lizards at a distance reveal influences of handling. *Copeia* 1988, 905-913.
- Rostro-García, S., Kamler, J. F., Crouthers, R., Sopheak, K., Prum, S., In, V., Pin, C., Caragiulo, A.,
  and Macdonald, D. W. (2018). An adaptable but threatened big cat: density, diet and prey selection of
  the Indochinese leopard (*Panthera pardus delacouri*) in eastern Cambodia. *Royal Society open science*5, 171-187.
- Rovero, F., Zimmermann, F., Berzi, D., and Meek, P. (2013). "Which camera trap type and how many
  do I need?" A review of camera features and study designs for a range of wildlife research applications. *Hystrix* 24, 148-156.
- 783

- Rowat, D., Speed, C. W., Meekan, M. G., Gore, M. A., and Bradshaw, C. J. (2009). Population
  abundance and apparent survival of the vulnerable whale shark, *Rhincodon typus*, in the Seychelles
  aggregation. *Oryx* 43, 591-598.
- 787
- Ruxmoore, R. and Groombridge, B. Asian Monitor Lizards: A Review of Distribution, Status,
  Exploitation, and Trade in Four Selected Species. 1990. (Secretariat of the Convention on International
  Trade in Endangered Species ....)
- Sanecki, G. M. and Green, K. (2005). A technique for using hair tubes beneath the snowpack to detect
  winter-active small mammals in the subnivean space. *European Journal of Wildlife Research* 51, 4147.
- 795

- Sannolo, M., Gatti, F., Mangiacotti, M., Scali, S., and Sacchi, R. (2016). Photo-identification in
   amphibian studies: a test of I3S Pattern. *Acta Herpetologica* 11, 63-68.
- 798

Schwarz, C. J. and Arnason, A. N. (1996). A general methodology for the analysis of capture-recapture
 experiments in open populations. *Biometrics* 52, 860-873.

801

Sears, M. W., Angilletta, M. J., Schuler, M. S., Borchert, J., Dilliplane, K. F., Stegman, M., Rusch, T.
W., and Mitchell, W. A. (2016). Configuration of the thermal landscape determines thermoregulatory
performance of ectotherms. *Proceedings of the National Academy of Sciences* 113, 10595-10600.

- Shine, R. (2010). The ecological impact of invasive cane toads (*Bufo marinus*) in Australia. *The Quarterly Review of Biology* 85, 253-291.
- 808

805

Shine, R. and Harlow, P. S. (1998). Ecological traits of commercially harvested water monitors, *Varanus salvator*, in northern Sumatra. *Wildlife research* 25, 437-447.

- 811812 Silver, S. C., Ostro, L. E., Marsh, L. K., Maffei, L., Noss, A. J., Kelly, M. J., Wallace, R. B., Gomez,
- H., and Ayala, G. (2004). The use of camera traps for estimating jaguar *Panthera onca* abundance and density using capture/recapture analysis. *Oryx* **38**, 148-154.
- 814 815

818

821

828

834

838

816 Smith, J., Brook, B., Griffiths, A., and Thompson, G. G. (2007). Can morphometrics predict sex in 817 varanids? *Journal of Herpetology* **41**, 133-141.

- Speed, C. W., Meekan, M. G., and Bradshaw, C. J. (2007). Spot the match–wildlife photo-identification
  using information theory. *Frontiers in zoology* 4.
- Stevenson, R. (1985). Body size and limits to the daily range of body temperature in terrestrial
  ectotherms. *The American Naturalist* 125, 102-117.
- Sutherland, D. R., Glen, A. S., and de Tores, P. J. (2010). Could controlling mammalian carnivores lead
  to mesopredator release of carnivorous reptiles? *Proceedings of the Royal Society B: Biological Sciences* 278, 641-648.
- Thompson, G. G. and Withers, P. C. (1997). Comparative morphology of western Australian varanid
  lizards (Squamata: *Varanidae*). *Journal of Morphology* 233, 127-152.
- Treilibs, C. E., Pavey, C. R., Hutchinson, M. N., and Bull, C. M. (2016). Photographic identification of
  individuals of a free-ranging, small terrestrial vertebrate. *Ecology and evolution* 6, 800-809.
- Tyne, J. A., Johnston, D. W., Christiansen, F., and Bejder, L. (2017). Temporally and spatially
  partitioned behaviours of spinner dolphins: implications for resilience to human disturbance. *Royal Society open science* 4, 160626.
- Volkov, I., Banavar, J. R., Hubbell, S. P., and Maritan, A. (2003). Neutral theory and relative species
  abundance in ecology. *Nature* 424, 1035-1037.
- Weinstein, B. G. (2018). A computer vision for animal ecology. *Journal of Animal Ecology* 87, 533545.
- 844
  845 Welbourne, D. (2013). A method for surveying diurnal terrestrial reptiles with passive infrared
  846 automatically triggered cameras. *PloS one* 6, e18965.
- 847
  848 Welbourne, D., Claridge, A., Paull, D., and Ford, F. (2019). Improving Terrestrial Squamate Surveys
  849 with Camera-Trap Programming and Hardware Modifications. *Animals* 9, 388.
- 850

- Welbourne, D. J., Claridge, A. W., Paull, D. J., and Lambert, A. (2016). How do passive infrared
  triggered camera traps operate and why does it matter? Breaking down common misconceptions. *Remote Sensing in Ecology and Conservation* 2, 77-83.
- 855 Kei 854
- Welbourne, D. J., MacGregor, C., Paull, D., and Lindenmayer, D. B. (2015). The effectiveness and cost
  of camera traps for surveying small reptiles and critical weight range mammals: a comparison with
  labour-intensive complementary methods. *Wildlife research* 42, 414-425.
- 858

- Welbourne, D. J., Paull, D. J., Claridge, A. W., and Ford, F. (2017). A frontier in the use of camera
  traps: surveying terrestrial squamate assemblages. *Remote Sensing in Ecology and Conservation* 3, 133145.
- Wickham, H. and Wickham, M. H. (2007). The ggplot package.)
- Wilson, S. K. and Swan, G. (2017) 'A complete guide to reptiles of Australia.' (Reed New HollandSydney.)
- 867
- 868 Withers, P. (2000). Overview of granite outcrops in Western Australia. Journal of the Royal Society of
- 869 *Western Australia* **83**, 103-108.
- 870 871